

# Unveiling causal relationship between non-covalent interactions and evaluated Young's modulus within oligolignols-cellulose complexes

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## Abstract

Linear correlation coefficients were calculated between the reported Young's modulus values and non-covalent interactions within cellulose-oligolignol complexes, considering the composition of an efficient adhesive formulation as previously reported. A paradigmatic relationship was observed. Molecular complexes of oligolignols with cellulose I $\beta$  were modeled using hybrid quantum mechanics/molecular mechanics (QM/MM) computations to obtain wavefunctions at the interaction region. Subsequently, a study of non-covalent interactions (NCI) based on the atoms in molecules (AIM) theory was implemented, utilizing graphics processing units (GPUs) for calculations. Our findings indicate that non-covalent interactions control the forces associated to adhesive-cellulose contacts, primarily through C-H $\cdots$ O hydrogen bonds, which promote the adhesion of oligolignols on cellulose I $\beta$ . Results indicate that the adhesion strength projected from larger YM values cannot be describe solely by the number of stronger hydrogen bonds nor by the number of the weak interactions but by the entire contributions of

specific interactions. Thus, significant linear correlations were observed between reported values Young's modulus and the molecular interactions observed, rendering the influence of oligolignol structure on the adhesion phenomenon in our cellulose I $\beta$  crystallite model. These observations promote the NCI and AIM analysis in a new framework to design adhesive formulations.

## Introduction

The need for eco-friendly wood adhesives has led to intense research on adhesives derived from natural and non-toxic materials (Pizzi, 2006, 2013; Antov, Savov & Neykov, 2020). Several studies have been reported on this topic, considering the lignin as one of the raw materials at the forefront of such studies (Nimz, 1983; Pizzi, 1994; Imam et al., 2001; Desai, Patel & Sinha, 2003; Li et al., 2004). On this context, the oligolignol-composition and Young's modulus (YM) experimental values from lignin-based adhesives, were postulated and identified by MALDI-TOF (matrix assisted laser desorption ionization time-of-flight) mass spectroscopy (Sedano-Mendoza, Lopez-Albarran & Pizzi, 2010a). In that study, a computational molecular docking analysis was employed to establish a correlation between the interaction energy of the proposed adhesives and a cellulose I $\beta$  crystallite model. These interaction energies were directly associated with the experimental YM values obtained through thermomechanical analysis (Martinez et al., 2010). The reported correlations in the study were considered acceptable. However, certain adhesive mixtures exhibited low correlation factors, despite their YM values implying effective adhesion. Naturally, the utilization of a molecular docking methodology serves as an initial approach to understand the interactions involved in the oligolignol-cellulose contact. Recently, we developed Reactive Force Field (ReaxFF) (Chenoweth, Van Duin & Goddard, 2008) molecular dynamic simulations to elucidate the contribution of each oligolignol's molecular structure to the adhesive capacity of the lignin mixture. We modelled the interaction of each oligolignol with a cellulose I $\beta$  crystallite model (López-Albarrán et al., 2017). The important results from those simulations were the coupling energies obtained for the oligolignol-cellulose complexes, which correlate quite well with experimental values of their Young's modulus. However, an *ab-initio* study is required to elucidate which molecular interactions are involved in the oligolignols-cellulose complexes, to determine which interactions address the adhesion effect beyond the energetic description. The intermolecular contacts between cellulose and adhesives constitute an important research area due to their potential in designing non-toxic glues for the wood industry (Fengel & Wegener, 1983; Houtman & Atalla, 1995; Imam et al., 2001; Desai, Patel & Sinha, 2003; Chenoweth, Van Duin & Goddard, 2008). Hydrogen bonding is undoubtedly a crucial topic for experimental and theoretical researchers (Nishiyama, Langan & Chanzy, 2002a; Ishikawa et al., 2015), as this interaction plays a key role in stabilizing the cellulose network (Moon et al., 2011; Djahedi et al., 2016). However, several authors (Cintrón, Johnson & French, 2011; Huang et al., 2021) have emphasized the need for careful consideration of both strong and weak hydrogen bonds, as these interactions offer insights into the design of materials with desired mechanical

properties. Unfortunately, *ab-initio* studies demand extensive computational time due to the large size of the systems involved.

In the composition of lignocellulosic biomass, lignin is a major component of wood, second only to cellulose, and acts as a cementing agent among wood cells.(Fengel & Wegener, 1983)

Extensive research from the scientific and engineering perspectives has been developed to investigate the interaction between cellulose and lignin, aiming to design and develop novel wood adhesive formulations (Pizzi, 1994; Pizzi & Mittal, 2011). The inclusion of lignin in these formulations has shown adhesive properties similar to those of phenol-formaldehyde resins (Jin, Cheng & Zheng, 2010; Pizzi, 2013), which find widespread application in industries involved in particleboard, plywood, fiberboard, and laminated wood (Navarrete et al., 2010; Mansouri et al., 2011)

Empirical studies have conducted on lignin-based adhesives (Knop & Scheib, 1979; Nimz, 1983). However, these studies have overlooked the molecular mechanism and the atomistic interactions within the cross-linked structure of lignin, specifically 4-hydroxyphenylpropanoids. Consequently, the driving forces behind the adhesion phenomenon remain unclear. Moreover, classical molecular dynamics simulations have been performed to investigate the absorption of lignin onto various cellulose surfaces (Besombes & Mazeau, 2005). Although the energy of interactions between lignin and cellulose has been reported, specific interactions relevant to adhesiveness, Young's modulus, or mechanical properties have not been addressed.

The lignin structure is primarily composed of simple units of 4-hydroxyphenylpropane, specifically three hydroxycinnamyl alcohols: p-coumarol, coniferol, and sinapol (Fig. 1). These units are commonly referred to as monolignols (Fengel & Wegener, 1983).

**Figure 1. Lignin precursors.** (A) p-coumarol, (B) coniferol, and (C) sinapol, all of them showed in their predominant (E)-monolignol configuration. Carbon numbering is shown on the coniferol structure.

In general, coniferol units are more abundant in softwood than in hardwood, as the latter exhibits a comparable proportion of coniferol and sinapol units (Shimada & Higuchi, 1983). The composition of lignin in solution differs from that of native lignin (protolignin) found in wood, owing to the extraction process (Terashima, Nakashima & Takabe, 1998). Despite these differences, the most abundant linkages observed between lignin units are as follows:  $\beta$ -O-4' ( $\beta$ O4),  $\beta$ -5' ( $\beta$ 5) and  $\beta$ - $\beta$ ' ( $\beta\beta$ ) (Sjöström, 1998).

In this work, we propose an atomistic analysis to investigate the role of the molecular structures of oligolignols in the adhesion properties of lignin-based formulations, as determined by Young's modulus. These structures have previously been characterized using the MALDI-TOF technique. Molecular dynamics simulations employing the ReaxFF are performed to model the coupling behavior of oligolignols on a cellulose I $\beta$  crystallite. Subsequently, a hybrid quantum mechanics/molecular mechanics (QM/MM) (Vreven & Morokuma, 2006) study is applied to

examine the obtained coupled molecular geometries of the oligolignol-cellulose complexes derived from the molecular dynamics study. Each complex is re-optimized to refine its conformation, which is then utilized to calculate its electron density ( $\rho(\mathbf{r})$ ) through density functional theory (DFT) (Kohn, Becke & Parr, 1996). The structures are subsequently analyzed using two tools developed within the quantum chemistry: 1) The non-covalent interaction index (NCI) (Johnson et al., 2010a), which explores regions where possible non-covalent interactions are present, and 2) the atoms in molecules (AIM) theory (Bader & Nguyen-Dang, 1981) which is a useful approach based on the electronic density ( $\rho(\mathbf{r})$ ) distribution within molecules in order to describe possible inter and intra non-covalent bonds. The NCI provides information on regions of non-covalent interactions, such as hydrogen bonds, weak interactions, and steric clashes, by evaluating the reduced gradient of the electron density (Boto et al., 2016), which is defined as:

$$s(\mathbf{r}) = \frac{1}{2(3\pi^2)^{\frac{1}{3}}} \frac{|\nabla\rho(\mathbf{r})|}{\rho(\mathbf{r})^{\frac{4}{3}}} \quad (1)$$

Therefore, the determination of electron density and its derivatives are essential for the NCI assessment. Furthermore, the identification of critical points ( $\mathbf{r}_c$ ) in the electron density,  $\rho(\mathbf{r}_c)$ , plays a crucial role in the AIM approach. Accordingly, the identification of  $\mathbf{r}_c$  satisfying the Laplacian  $\nabla^2\rho(\mathbf{r}_c) = 0$  is important for the analysis of the electron density as these points indicate bond paths and bond critical points. Thus, these quantum chemistry tools serve to describe and elucidate both short- and long-range interactions involved in the adhesion of oligolignols to the cellulose I $\beta$  surface. Evidently, the aim of this work is to establish a foundation for designing and developing adhesive formulations for lignocellulosic materials by gaining insight into their fundamental molecular interactions with a cellulose I $\beta$  surface. As far as we know, no previous theoretical reports have employing reactive molecular dynamics in conjunction with NCI-AIM methodologies to elucidate the interaction between oligolignols and cellulose. This research represents a key step towards the development of new and eco-friendly adhesive formulations for wood.

Naturally, performing NCI-AIM evaluations along with their corresponding derivatives can be a challenging when dealing with considerable systems sizes. To address this issue, we employ the GPUAM code, a QM program specifically designed to handle large systems and optimized for graphics processing units (GPUs) (Hernández-Esparza et al., 2014). GPUAM has demonstrated its capability to deliver reliable results in less computing time compared to software developed for x86 multicore CPUs (Hernández-Esparza et al., 2014).

We believe that this type of study provides a framework for elucidating oligolignols or lignin substructures within lignin-adhesive formulations that augment cellulose adhesion. Besides, the present study can contribute to delineating the interactions between lignin components and cellulose.

## Materials & Methods

Considering that the most prevalent form of the cellulose in higher plants is cellulose I $\beta$  (Nishiyama, Langan & Chanzy, 2002b), the cellulose model used in this work was constructed based on crystalline structures characterized through X-ray and neutron fiber diffraction measurements (Nishiyama, Langan & Chanzy, 2002a; Nishiyama et al., 2003). Our cellulose I $\beta$  crystallite model comprises three layers in the  $y$  direction and six glucan chains in the  $z$  direction. Each chain consists of 12 glucose units (equivalent to six cellobioses) in the  $x$  direction, as illustrated in Fig. 2.

**Figure 2. Snapshot of the cellulose I $\beta$  model used to form the complexes with the oligolignols.** The model was built using the Cellulose-Builder toolkit (Gomes & Skaf, 2012), and visualized by the Vesta molecular viewer (Momma & Izumi, 2008). Color key: carbons atoms are in dark-gray, oxygen atoms in red, and hydrogen atoms in white.

The molecular structures of oligolignols elucidated by our MALDI-TOF experimental results, were constructed using the ChemBioOffice Ultra software version 11.0.1 (Ultra, 2007). These oligolignols were assumed to interact with the cellulose I $\beta$  crystallite model. The set of oligolignols examined in this study (see Fig. 3) was characterized from the experimental adhesive formulation with the best performance (highest value for Young's modulus) (Pizzi et al., 2011). As observed, the modeled oligolignols are constituted only by coniferyl (Fig. 1B) and sinapyl (Fig. 1C) units. We employed the following nomenclature for oligolignols: A\_m\_B\_n\_C where A, B or C indicates a coniferyl (CA) or sinapyl (SA) alcohol linked by one  $m$  or  $n$  linkage (for instance,  $\beta$ O4,  $\beta\beta$ , and  $\beta$ 5). Therefore, we constructed the following oligolignols CA\_ $\beta$ O4\_SA\_ $\beta$ 5\_CA (Fig. 3A), CA\_ $\beta$ O4\_SA\_ $\beta\beta$ \_SA (Fig. 3B), SA\_ $\beta$ O4\_SA\_ $\beta\beta$ \_SA (Fig. 3C), and CA\_ $\beta\beta$ \_CA (Fig. 3D).

**Figure 3. Molecular structures of the oligolignols in the reported efficient adhesive formulation.** (A) CA\_ $\beta$ O4\_SA\_ $\beta$ 5\_CA, (B) CA\_ $\beta$ O4\_SA\_ $\beta\beta$ \_SA, (C) SA\_ $\beta$ O4\_SA\_ $\beta\beta$ \_SA, and (D) CA\_ $\beta\beta$ \_CA.

The present Quantum Mechanics (QM)/Molecular Mechanics (MM) study utilized the four oligolignol-cellulose complexes previously reported in the most effective oligolignol-mixture (Sedano-Mendoza, Lopez-Albarran & Pizzi, 2010b; López-Albarrán et al., 2017). The study employed the ONIOM method (Dapprich et al., 1999), implemented in the Gaussian16 suite code (Frisch et al., 2016). This methodology enables the analysis of the primary interactions using an *ab-initio* approach. In our proposed approach, the QM region comprised the oligolignol and a cellulose I $\beta$  surface fragment, while the remainder of the cellulose model was designated as the MM region. The four oligolignol-cellulose complexes were geometrically minimized applying the ONIOM method which defines two or three layers (molecular or atomic) within the

structure that are evaluated at different levels of molecular modeling. The geometry optimization for each complex, was performed including long range interactions by using the CAM-B3LYP (Yanai, Tew & Handy, 2004) method and the cc-pVDZ Dunning (Kendall, Dunning Jr & Harrison, 1992) basis set. The MM contribution was calculated applying the universal force field (UFF) (Rappé et al., 1992) implemented in the Gaussian16 suit package. The output coordinates obtained from the QM/MM minimization process were employed to compute the electronic wavefunction via a single point energy calculation for the QM region (oligolignol and part of the cellulose complex). These calculations were carried out considering that the QM region requires an *ab-initio* method or methods developed to describe van der Waals (vdW) interactions. In this work, the M06-2X functional (Zhao & Truhlar, 2008), including dispersion interactions proposed by Grimme in the commonly form named “D3”(Grimme, 2004), was used for vdW corrections. Recently, different Kohn–Sham approaches have been employed to test the behavior of some cellulose properties by including dispersion interactions in periodic systems (Navarrete-López, San-Román & Zicovich-Wilson, 2016). The resulting wavefunction was then analyzed to evaluate the NCI interactions associated with the AIM approach for each system. The evaluation of NCI index and the bond critical points related to AIM theory, were obtained by using a grid-based method. This grid-based method was developed and implemented by Hernández-Esparza et al. on graphics processing units (GPUs), which is quite convenient to reduce considerably the computation efforts, particularly in large systems, such as the oligolignol-cellulose complexes studied in this manuscript. Subsequently, the observed interactions were classified into  ${}_{\text{adh}}\text{OH}\cdots\text{O}_{\text{Cell}}$ ,  ${}_{\text{adh}}\text{O}\cdots\text{HO}_{\text{Cell}}$ ,  ${}_{\text{adh}}\text{CH}\cdots\text{O}_{\text{Cell}}$ ,  ${}_{\text{adh}}\text{O}\cdots\text{HC}_{\text{Cell}}$ ,  ${}_{\text{adh}}\text{C}\pi\text{H}\cdots\text{O}_{\text{Cell}}$ , and  ${}_{\text{adh}}\text{C}\pi\cdots\text{HC}_{\text{Cell}}$ , based on its electron density and the Laplacian of the electron density (which must be positive for all cases). Among them, weak interactions were considered as all of interactions with electron density value less than 0.005 a.u. (Johnson et al., 2010a). Finally, a series of correlations between the observed interactions and the previously reported experimental values for the Young’s modulus were determined.

## Results

The molecular structures of the oligolignol-cellulose complexes optimized at the interaction region through the QM/MM-ONIOM studies, remain almost in the same conformation than those obtained from our previous LAMMPS-ReaxFF-MD study (López-Albarrán et al., 2017). The superposition of the obtained oligolignols conformations in this QM/MM study against those reported from our LAMMPS-ReaxFF-MD simulations, is displayed in Fig. 4. The cellulose model was not displayed, to facilitate visualization; however, the entire system is displayed in Fig. S1. Additionally, the QM and MM regions for each oligolignol-cellulose complexes are displayed in the Fig. S2 in the Supplemental Information.

**Figure 4. Superposition of the four oligolignols complexed over the cellulose Iβ crystallite model.** For convenience the cellulose Iβ surface is not present in this figure however, this part of

the system was considered in the QM/MM optimization process. The complex conformations obtained from the LAMMPS-ReaxFF-MD are in green carbon-skeleton, and the QM/MM-CAM-B3LYP(Yanai, Tew & Handy, 2004)/cc-pVDZ(Kendall, Dunning Jr & Harrison, 1992)/Universal Force Field(Rappé et al., 1992) (UFF) are in purple carbon-skeleton. (A) CA\_βO4\_SA\_β5\_CA trilignol, (B) CA\_βO4\_SA\_ββ\_SA trilignol, (C) SA\_βO4\_SA\_ββ\_SA trilignol, and (D) CA\_ββ\_CA dilignol.

The molecular interactions arising from the adhesion between the cellulose Iβ crystallite model and each oligolignol were analyzed using NCI-AIM surfaces to establish relationships between these interactions and the Young's modulus. The intermolecular interactions are presented in Fig. 5, with only QM regions displayed. The localization of the NCIs through the surfaces was related to the reduced gradient density (RGD) plot regions, similar to the approach by Ponnuchamy, Sandak & Sandak, (2020). Due to the absence of steric clashes (red isosurfaces), adhesion is promoted by attractive non-covalent interactions (green and blue isosurfaces). In Fig. 5A, the CA\_βO4\_SA\_β5\_CA-cellulose Iβ complex shows strong interactions (blue isosurfaces) labeled as **a-d**. The system also displays relative weak interactions with electron density values around 0.011 a.u. (labels **e-j**). Additionally, there are large green isosurfaces with small electron density values (<0.002 a.u., labeled as **l** and **m**). Strong hydrogen bonds labeled as **a** and **b** are found in the CA\_βO4\_SA\_ββ\_SA-cellulose Iβ complex (Fig. 5B). Moreover, localized contacts (small green isosurfaces related two atoms) are indicated with **c-f**, **j**, **m**, **n** and **q-s** labels. These interactions span a range from ~0.001 a.u to ~0.015 a.u. In the case of SA\_βO4\_SA\_ββ\_SA-cellulose Iβ complex in Fig. 5C, one strong hydrogen bond is noticeable, as indicated by the **a** label. Localized contacts are indicated with **b-d**, **f**, **g**, **k**, **q**, **s**, **t**, and **u** labels in the range ~0.001 a.u. to 0.012 a.u. Interestingly, this number of interactions is similar but weaker than those found in Complex 2. Finally, for the CA\_ββ\_CA-cellulose Iβ (Fig. 5D), despite the lower number of interactions compared to previous systems, the localized **b**, **c**, and **e** contacts extend over a narrower but stronger 0.003 a.u. to 0.012 a.u. range compared to the remaining complexes.

**Figure 5. Mapping of the NCI-AIM surfaces and plot of reduced electron density gradient (RDG) against  $\text{Sign}(\lambda_2)\rho$ .** (A) Complex 1, CA\_βO4\_SA\_β5\_CA– cellulose Iβ; (B) Complex 2, CA\_βO4\_SA\_ββ\_SA–cellulose Iβ; (C) Complex 3, SA\_βO4\_SA\_ββ\_SA–cellulose Iβ; (D) Complex 4, CA\_ββ\_CA–cellulose Iβ. Blue regions are for hydrogen bonds, green regions for van der Waals. The gradient isosurfaces correspond to a cutoff of 0.5 a.u. Color scale ranging from  $-0.4 < \rho < 0.4$  a.u. The interactions labeled as **a-u** (if exist) corresponded to the interactions indicated in Tables S1-S4.

Additionally, the AIM approach was used to determine possible (localized) contacts between atoms from oligolignols and atoms from the cellulose surface, through bond paths. These bond



paths are displayed in Fig. 6 for each oligolignol-cellulose complex. By analyzing bond paths, a series of hydrogen bonds: O-H $\cdots$ O, C-H $\cdots$ O and C $\pi$ -H $\cdots$ O were found. These hydrogen bonds presented electronic density values between 0.006 and 0.040 a.u., accordingly to the electronic density criterion from Johnson et al.(Johnson et al., 2010a) A detailed analysis of these contacts, including weak interactions, is presented in Tables S1 to S4 in the Electronic Supplemental Information.

**Figure 6. Illustration of the main hydrogen bond based on electronic density isovalues and a blue code color:  $\rho(r) \geq 0.02$  in dark blue,  $0.01 \leq \rho(r) < 0.02$  in blue and  $\rho(r) < 0.01$  in cyan.**(Johnson et al., 2010b) Atoms involved in hydrogen bond are highlighted in magenta. (A) Complex 1, CA\_βO4\_SA\_β5\_CA – cellulose Iβ; (B) Complex 2, CA\_βO4\_SA\_ββ\_SA – cellulose Iβ; (C) Complex 3, SA\_βO4\_SA\_ββ\_SA – cellulose Iβ; (D) Complex 4, CA\_ββ\_CA – cellulose Iβ.

## Discussion

The adhesion process for each complex is driven by non-covalent interactions, as evidenced by the occurrence of both small and large green zones between oligolignol and cellulose fragments with few blue zones (typically labeled as **a-d** in Fig. 5), indicative of strong hydrogen bonds (Johnson et al., 2010a). Although these interactions may appear covalent at first glance due to strong binding, like previous reports where strong hydrogen bonds stabilize DNA base pairs (Jurečka et al., 2006), it is important to note that they are, in fact, non-covalent in nature. The significance of  $\rho(r)$  values is acknowledged, with larger values corresponding to greater binding energies for hydrogen bonds (Koch & Popelier, 1995; Navarrete-López, Garza & Vargas, 2007). However, it is not easy to know the contribution of each contact to the binding energy. Naturally, the possible cooperativity exhibited by hydrogen bonds in these systems is one important topic to be analyzed in future studies. In addition, the many-body problem involved in the lignin-cellulose interaction is one important factor that is not addressed in this report and our conclusions are limited by the QM/MM model. For that reason, it is not possible to compare one to one binding energies from QM with ReaxFF-MD. Notwithstanding these limitations, the non-covalent interaction (NCI) interactions depicted in Fig. 5, and the bond paths reported in Fig.6 offer valuable insights. The examination of these interactions may unveil potential connections between the macroscopic Young's modulus values and the specific punctual interactions arising after the contact between cellulose and oligolignol. In this context, the study aims to elucidate the atomistic interactions or their interconnections governing the adhesion phenomenon through YM. Such interactions can validate the influence of oligolignols composition and contribute to the synthesis of efficient adhesive formulations. The reported YM values among the four oligolignols from the best tested depolymerized lignin, shown the following order for the optimal adhesion between oligolignols and cellulose Iβ:

CA\_βO4\_SA\_ββ\_SA(0.3011 MPa)>CA\_ββ\_CA(0.2988 MPa)>SA\_βO4\_SA\_ββ\_SA(0.2093 MPa)>CA\_βO4\_SA\_β5\_CA(0.1346 MPa) (Sedano-Mendoza, Lopez-Albarran & Pizzi, 2010b). The trilignol with the lowest YM value exhibited inefficient adhesion, as elucidated by our NCI-AIM study. The CA\_βO4\_SA\_β5\_CA trilignol did not achieved optimal coupling over the cellulose Iβ crystallite model, yielding complexes that do not promote adhesion. One reason may be that the trilignol exhibits folding (see Fig. 5A), reducing intermolecular non-covalent interactions. This conformation produces large green isosurfaces related to non-localized contacts. Interestingly, this oligolignol present the strongest hydrogen bonds (**a** and **b** interactions located at electron density values of 0.04 and 0.038 a.u., respectively) but also the lowest YM value (0.1346 MPa). In contrast, the CA\_βO4\_SA\_ββ\_SA trilignol shows optimal coupling on the cellulose Iβ crystallite, reducing the large isosurfaces and increasing the localized non-covalent interactions (**a-s**), as shown in Fig. 5B. This combined effect promotes oligolignol adhesion due to the occurrence of several both hydrogen bonds and weak interactions towards the cellulose crystallite, see Table S2. It is noteworthy that, despite producing the largest YM value, the CA\_βO4\_SA\_ββ\_SA trilignol has weaker hydrogen bonds and a large number of weak interactions compared to those found in Complex 1. Similar coupling is observed for the SA\_βO4\_SA\_ββ\_SA–cellulose Iβ complex, shown in Fig. 5C. However, it presents weaker hydrogen bonds and a small number of weak interactions (see Table S3). Finally, the CA\_ββ\_CA dilignol achieves efficient adhesion due to its large YM value (Sedano-Mendoza, Lopez-Albarran & Pizzi, 2010b). From the NCI-AIM analysis, it was found that this dilignol displays a relatively low number of weak non-covalent interactions; however, they are concentrated in a narrow range, as can be seen from Table S4 and indicated previously. Moreover, the dilignol did not show strong hydrogen bonds, as the plot of reduced electron density gradient (RDG) against  $\text{Sign}(\lambda_2)\rho$  suggests, shown in Fig. 5D. However, experimental and theoretical results from the literature support that the CA\_ββ\_CA oligolignol is the second-best lignin substructure contributing to the adhesion (Mansouri et al., 2011), as it was the unique substructure plausible to couple more dilignols over the cellulose surface. The previous analysis suggests that both weak non-covalent interactions and hydrogen bonds likely play a complementary role in mechanical properties such as Young's modulus. To explore this effect further, the reported interactions were classified, and their respective electron density values were further analyzed. The number and type of molecular interactions between the cellulose Iβ crystallite and each oligolignol-adhesive, was reported through the histogram shown in Fig. 7, under the following descriptions: blue columns correspond to the CA\_βO4\_SA\_β5\_CA trilignol (Complex 1), green columns for CA\_βO4\_SA\_ββ\_SA trilignol (Complex 2), yellow columns for SA\_βO4\_SA\_ββ\_SA trilignol (Complex 3), and red columns CA\_ββ\_CA dilignol (Complex 4). The non-covalent interactions found in each complex were classified as indicated above, i.e.,  $\text{adhOH}\cdots\text{O}_{\text{Cell}}$ ,  $\text{adhO}\cdots\text{HO}_{\text{Cell}}$ ,  $\text{adhCH}\cdots\text{O}_{\text{Cell}}$ ,  $\text{adhO}\cdots\text{HC}_{\text{Cell}}$ , and  $\text{adhC}\pi\cdots\text{HC}_{\text{Cell}}$  (or  $\text{adhC}\pi\text{H}\cdots\text{O}_{\text{Cell}}$ ).

**Figure 7. Summary of non-covalent interactions found in the four oligolignol-cellulose complexes.**

Complexes 02 and 03 presented the highest number of interactions, with hydrogen bonds of the type “ ${}_{\text{adh}}\text{CH}\cdots\text{O}_{\text{Cell}}$ ” predominating for Complex 3, while “ ${}_{\text{adh}}\text{O}\cdots\text{HC}_{\text{Cell}}$ ” hydrogen bonds were more prevalent in Complex 2. Despite that Complex 2 (CA- $\beta$ O4-SA- $\beta$  $\beta$ -SA-cellulose I $\beta$ ) shows only three interactions namely  $\text{O}\cdots\text{HO}$ , it also has a larger number of  $\text{CH}\cdots\text{O}$  interactions, which can be associated with the largest YM value (Sedano-Mendoza, Lopez-Albarran & Pizzi, 2010b). Indeed, in this complex, a larger localized distribution of non-covalent interactions was found, while in Complex 3, predominant methyl hydrogen bonds were observed. From a molecular level description, this result elucidates which oligolignol promotes adhesion over cellulose, revealing the atomistic interactions involved in the oligolignol-cellulose complex based on the NCI-AIM study. The oligolignol CA- $\beta$  $\beta$ -CA occurred in Complex 4, where it couples a guaiacyl fragment on the cellulose in a coplanar configuration, yielding closer intermolecular non-covalent interactions per dilignol coupled on the cellulose. In other words, weak interactions with density values larger than 0.003 a.u. are observed in this complex (Table S4). Interestingly, Complex 4 is reported as the second-best lignin substructure contributing to the adhesion.

Clearly, the  $\text{CH}\cdots\text{O}$  interaction predominates over  $\text{OH}\cdots\text{O}$  and  $\text{C}\pi\text{H}\cdots\text{O}$  interactions, as displayed in Table S4. Thus, weak interactions like the  $\text{CH}\cdots\text{O}$  contact hold significant relevance in binding oligolignols with cellulose, despite the fact that the larger interaction present only has a value of 0.023 a.u.; a phenomenon previously noted in analogous systems (Vargas et al., 2000a,b; Salazar-Cano et al., 2016). This aligns with the findings of several authors (Cintrón, Johnson & French, 2011; Huang et al., 2021) who suggest the important role of weak interactions over strong hydrogen bonds.

As mentioned earlier, neither stronger hydrogen bonds nor the quantity of weak interactions can be directly related to experimental values of the Young's modulus. Therefore, there seems to be a reciprocal influence between such interactions at the coupling of oligolignol. For instance, a substantial presence of strong hydrogen bonds, as observed in Complex 1, may give rise to regions with non-localized interactions. Conversely, a significant number of localized yet predominantly weak interactions, exemplified by Complex 3, might lead to a fragile support structure for oligolignol. In this context, the impact of each non-covalent interaction on the adhesion of oligolignols to cellulose was assessed by establishing causal relationships with the experimental YM values. Firstly, we plotted the average electronic density resulting from the  ${}_{\text{adh}}\text{CH}\cdots\text{O}_{\text{Cell}}$  interaction against the experimental YM values (refer to Fig. 8), achieving a correlation factor of  $R^2 = 0.97$ .

**Figure 8. Correlation between the average electron density of the  ${}_{\text{adh}}\text{CH}\cdots\text{O}_{\text{Cell}}$  interactions and Young's modulus for the evaluated complexes.**

In Fig. 8, it is evident that the larger the average electron density of the  $_{\text{adh}}\text{CH}\cdots\text{O}_{\text{Cell}}$ , the lower the Young's modulus. Interestingly, the  $_{\text{adh}}\text{CH}\cdots\text{O}_{\text{Cell}}$  interactions observed in Complex 1 and Complex 3 result in larger values of the  $\alpha$  angle (Table S1 and Table S3). Consequently, the adhesive components must adjust their positions to establish the particular  $_{\text{adh}}\text{CH}\cdots\text{O}_{\text{Cell}}$  interaction, which could promote the formation of other crucial interactions. Furthermore, by plotting the total contribution of the  $_{\text{adh}}\text{XH}\cdots\text{O}_{\text{Cell}}$  interactions (with X as C, C $\pi$  and O atoms from oligolignols), against YM values, a linear correlation was also found ( $R^2 = 0.79$ ), as depicted in Fig. 9. Interestingly, the slope of the aforementioned linear correlations was negative, indicating that such interactions limit the stretchability of the adhesive or achieves a rigid state, as suggested by Huang et al. (Huang et al., 2021) in the evaluation of mechanical properties of polymers.

**Figure 9. Correlation obtained between the total electron density contribution of  $_{\text{adh}}\text{XH}\cdots\text{O}_{\text{Cell}}$  interactions and Young's modulus for the evaluated complexes.**

In contrast, plotting the total contribution of the weak interactions (interactions with electron density values below 0.005 a.u.) against the YM values yielded a positive slope and a correlation factor of  $R^2 = 0.82$ , as shown in Fig. 10. This significant correlation suggests that weak interactions play a fundamental role in enhancing adhesion by providing stabilization along the complex. Surprisingly, these linear correlations represent the first report to render the effect of non-covalent interactions on the mechanical parameter YM.

**Figure 10. Correlation obtained between the total electron density contribution of weak interactions ( $\rho \leq 0.005$  a.u.) and Young's modulus for the evaluated complexes.**

Finally, by evaluating the ratio of the total contribution from  $_{\text{adh}}\text{XH}\cdots\text{O}_{\text{Cell}}$  to the total contribution of weak interactions, a significant correlation ( $R^2 = 0.87$ ) was also found, as shown in Fig. 11. This correlation, with a negative slope, suggests that better oligolignol adhesives can be obtained if both contributions are comparable, in agreement with the observations from Fig. 10. Accordingly, hydrogen bonds and weak interactions must play a complementary role in synthesizing better and environmentally friendly lignin-based adhesives.

**Figure 11. Correlation obtained between the ratio of total electron density contribution of  $_{\text{adh}}\text{XH}\cdots\text{O}_{\text{Cell}}$  and weak interactions and Young's modulus for the evaluated complexes.**

It is noteworthy that the adhesion between cellulose and the best depolymerized lignin adhesive-formulation, can be elucidate from molecular modeling studies, considering an atomistic point of view where only one molecule of oligolignol interacts with the cellulose model. However, the experimental data were considered in assays where the solvents (mainly water) were removed by heating until the lignin adhesive formulations reached their Young's modulus.

## Conclusions

Based on the established correlation between molecular dynamic interaction energy and Young's modulus (López-Albarrán et al., 2017), the triligol CA\_βO4\_SA\_ββ\_SA and the dilignol CA\_ββ\_CA were identified as the lignin substructures contributing to the adhesion. These oligolignols exhibited favorable non-covalent interactions in this NCI-AIM study. Consequently, the adhesion between lignin-based adhesives and cellulose can be screened from specific non-covalent interactions between oligolignols and the cellulose model.

The coupling of oligolignols such as CA\_βO4\_SA\_β5\_CA on the cellulose Iβ surface induces the folding of the triligol, resulting in extensive green (weak) isosurfaces associated with non-localized interactions. This contrasts with the CA\_ββ\_CA-cellulose Iβ complex.

Deciphering the precise contribution of each non-covalent interaction to the binding energy is challenging. Nevertheless, to provide insights into this aspect, the roles of various oligolignol-cellulose interactions were described. Notably, C-H...O contacts were prevalent, with oligolignols serving as hydrogen atom donors in some instances and cellulose acting as the donor in others. Furthermore, weak interactions such as the CH...O contact played a significant role in binding oligolignols with cellulose. Examining the electron density of the H-bonds  $_{adh}XH \cdots O_{Cell}$  and weak interactions revealed their complementary roles in fully describing the adhesive phenomenon. A paradigmatic relationship was observed, the linear correlations established in this study represent the first report demonstrating the influence of non-covalent interactions on the measurable mechanical parameter YM. Our findings indicate that non-covalent interactions control the forces associated to adhesive-cellulose contacts, primarily through C-H...O hydrogen bonds, which promote the adhesion of oligolignols on cellulose Iβ. Results indicate that the adhesion strength projected from larger YM values cannot be describe solely by the number of stronger hydrogen bonds nor by the number of the weak interactions but by the entire contributions of specific interactions.

## Additional Information and Declarations

The following information was supplied regarding data availability: The interaction correlations from raw data and the scripting code for figures are available in the Supplemental Files Data S1-S6.

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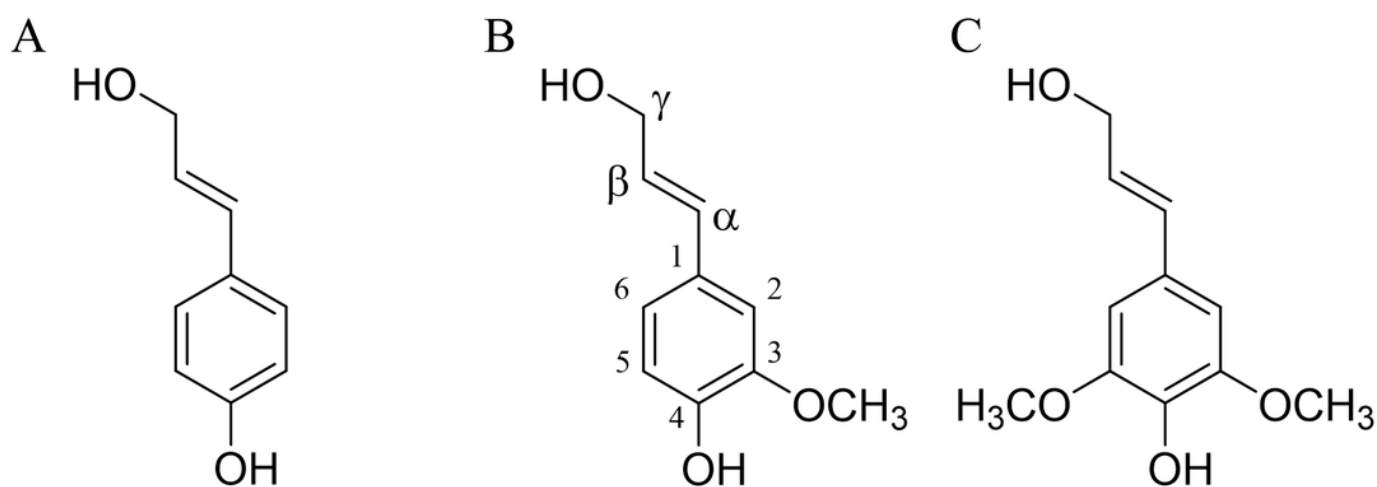


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# Figure 1

Lignin precursors.

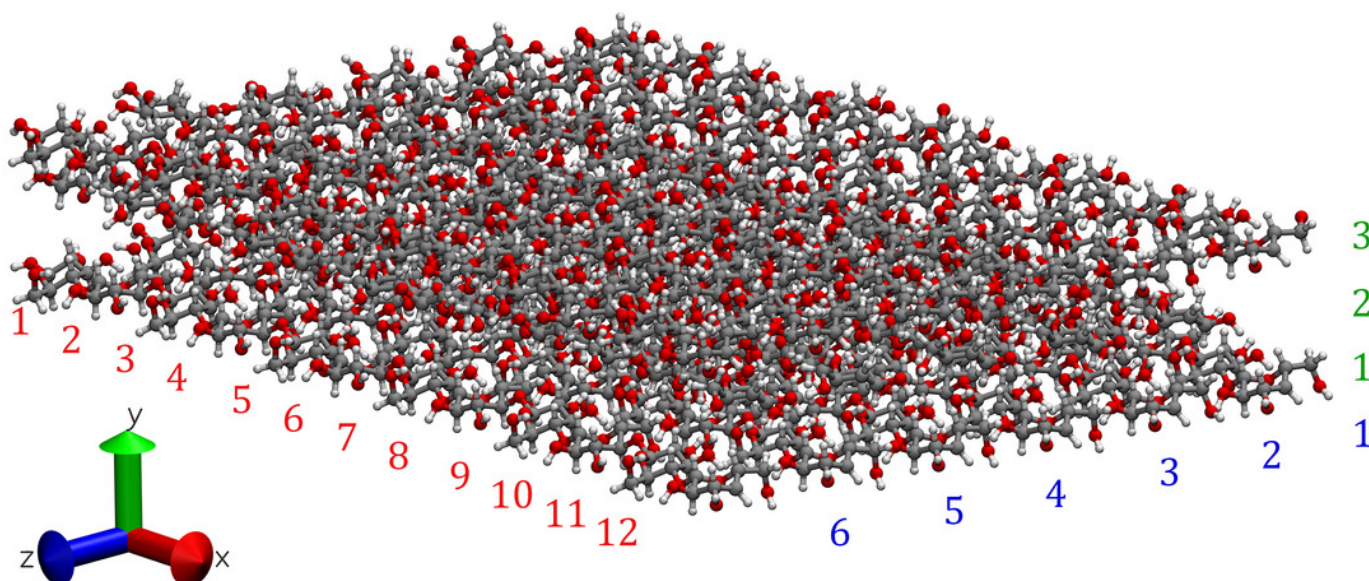
**Lignin precursors.** (A) p-coumarol, (B) coniferol, and (C) sinapol, all of them showed in their predominant (E)-monolignol configuration. Carbon numbering is shown on the coniferol structure.



## Figure 2

Snapshot of the cellulose I $\beta$  model used to form the complexes with the oligolignols.

The model was built using the Cellulose-Builder toolkit,<sup>(Gomes & Skaf, 2012)</sup> and visualized by the Vesta molecular viewer.<sup>(Momma & Izumi, 2008)</sup> Color key: carbons atoms are in dark-gray, oxygen atoms in red, and hydrogen atoms in white.

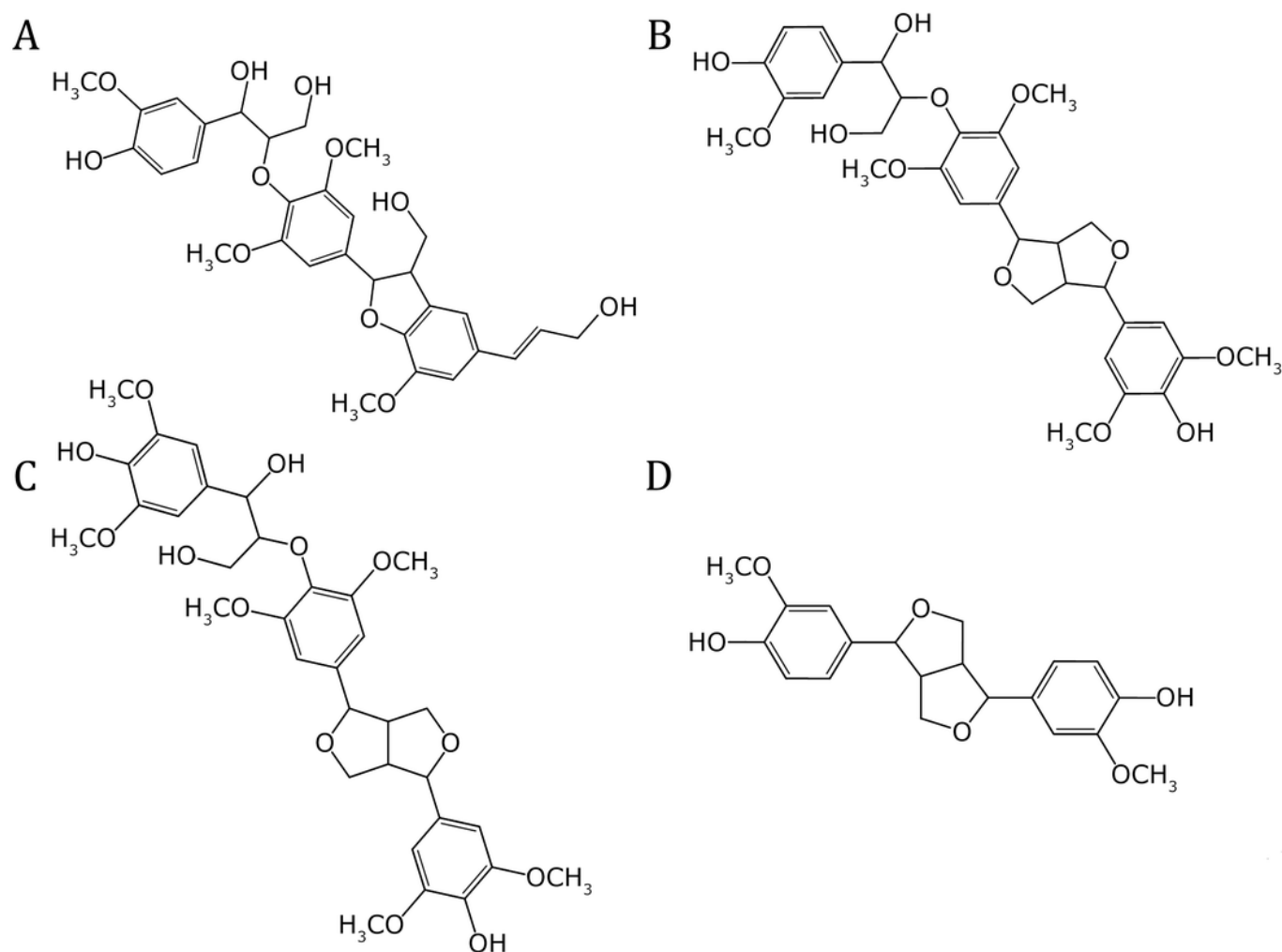


## Figure 3

Molecular structures of the oligolignols

**Molecular structures of the oligolignols in the reported efficient adhesive**

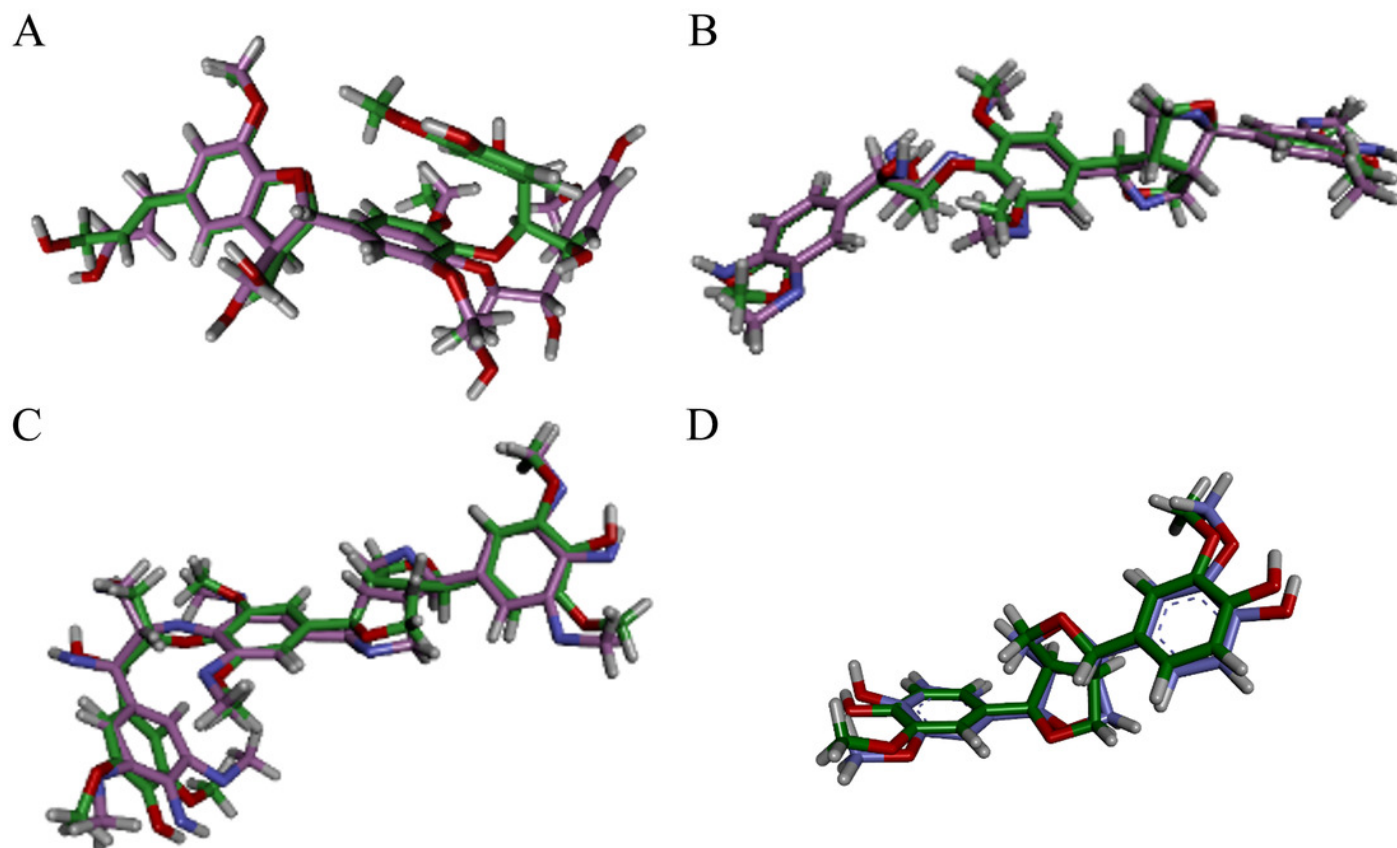
**formulation.** (A) CA- $\beta$ O4-SA- $\beta$ 5-CA, (B) CA- $\beta$ O4-SA- $\beta\beta$ -SA, (C) SA- $\beta$ O4-SA- $\beta\beta$ -SA, and (D) CA- $\beta\beta$ -CA.



## Figure 4

Superposition of the four oligolignols complexed over the cellulose I $\beta$  crystallite model.

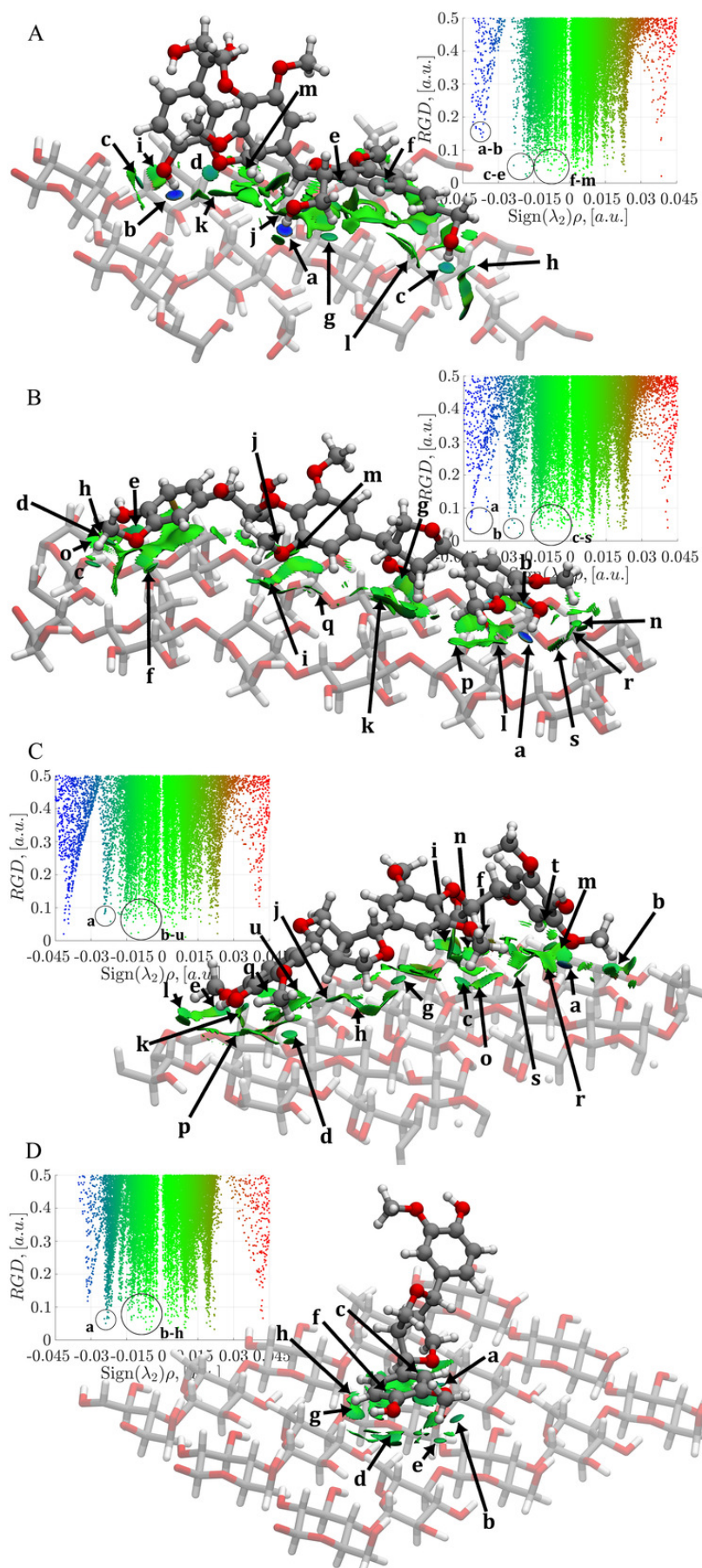
**Superposition of the four oligolignols complexed over the cellulose I $\beta$  crystallite model.** For convenience the cellulose I $\beta$  surface is not present in this figure however, this part of the system was considered in the QM/MM optimization process. The complex conformations obtained from the LAMMPS-ReaxFF-MD are in green carbon-skeleton, and the QM/MM-CAM-B3LYP<sup>(Yanai, Tew & Handy, 2004)</sup>/cc-pVDZ<sup>(Kendall, Dunning Jr & Harrison, 1992)</sup>/Universal Force Field (Rappé et al., 1992) (UFF) are in purple carbon-skeleton. (A) CA $\beta$ O4\_SA $\beta$ 5\_CA trilignol, (B) CA $\beta$ O4\_SA $\beta\beta$ \_SA trilignol, (C) SA $\beta$ O4\_SA $\beta\beta$ \_SA trilignol, and (D) CA $\beta\beta$ \_CA dilignol.



## Figure 5

Mapping of the NCI-AIM surfaces and plot of reduced electron density gradient (RDG) against  $\text{Sign}(\lambda_2)$

**Mapping of the NCI-AIM surfaces and plot of reduced electron density gradient (RDG) against  $\text{Sign}(\lambda_2)$ .** (A) Complex 1, CA\_βO4\_SA\_β5\_CA-cellulose Iβ; (B) Complex 2, CA\_βO4\_SA\_ββ\_SA-cellulose Iβ; (C) Complex 3, SA\_βO4\_SA\_ββ\_SA-cellulose Iβ; (D) Complex 4, CA\_ββ\_CA-cellulose Iβ. Blue regions are for hydrogen bonds, green regions for van der Waals. The gradient isosurfaces correspond to a cutoff of 0.5 a.u. Color scale ranging from  $-0.02$  a.u. to  $0.02$  a.u. The interactions labeled as **a-u** (if exist) corresponded to the interactions indicated in Tables S1-S4.



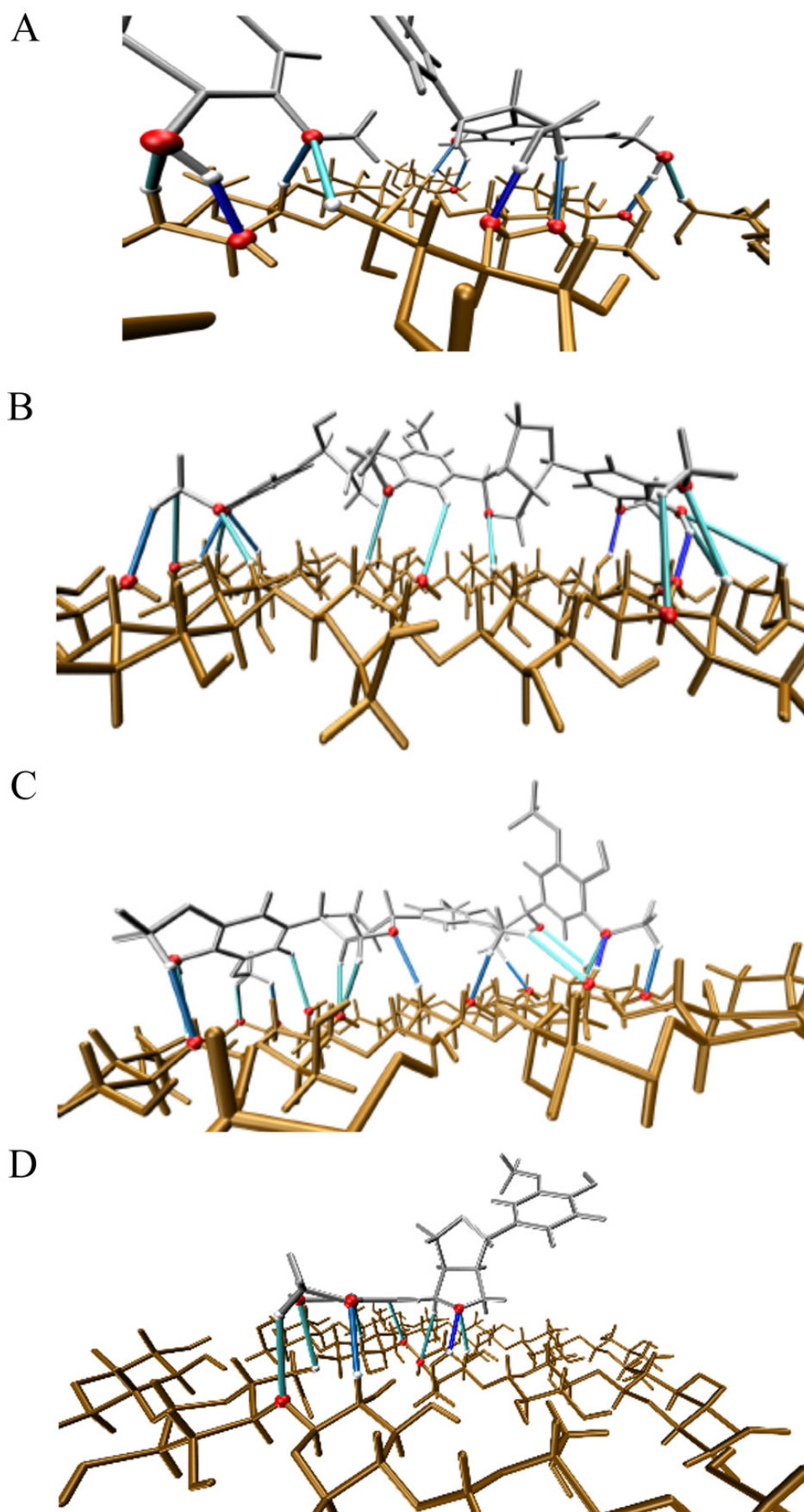


## Figure 6

Illustration of the main hydrogen bond based on electronic density isovalues

Illustration of the main hydrogen bond based on electronic density isovalues and a blue code color:  $\rho(r) \geq 0.02$  in dark blue,  $0.01 \leq \rho(r) < 0.02$  in blue and  $\rho(r) < 0.01$  in cyan. (Johnson et al., 2010b) Atoms involved in hydrogen bond are highlighted in magenta. (A) Complex 1, CA\_βO4\_SA\_β5\_CA - cellulose Iβ; (B) Complex 2, CA\_βO4\_SA\_ββ\_SA - cellulose Iβ; (C) Complex 3, SA\_βO4\_SA\_ββ\_SA - cellulose Iβ; (D) Complex 4, CA\_ββ\_CA - cellulose Iβ.

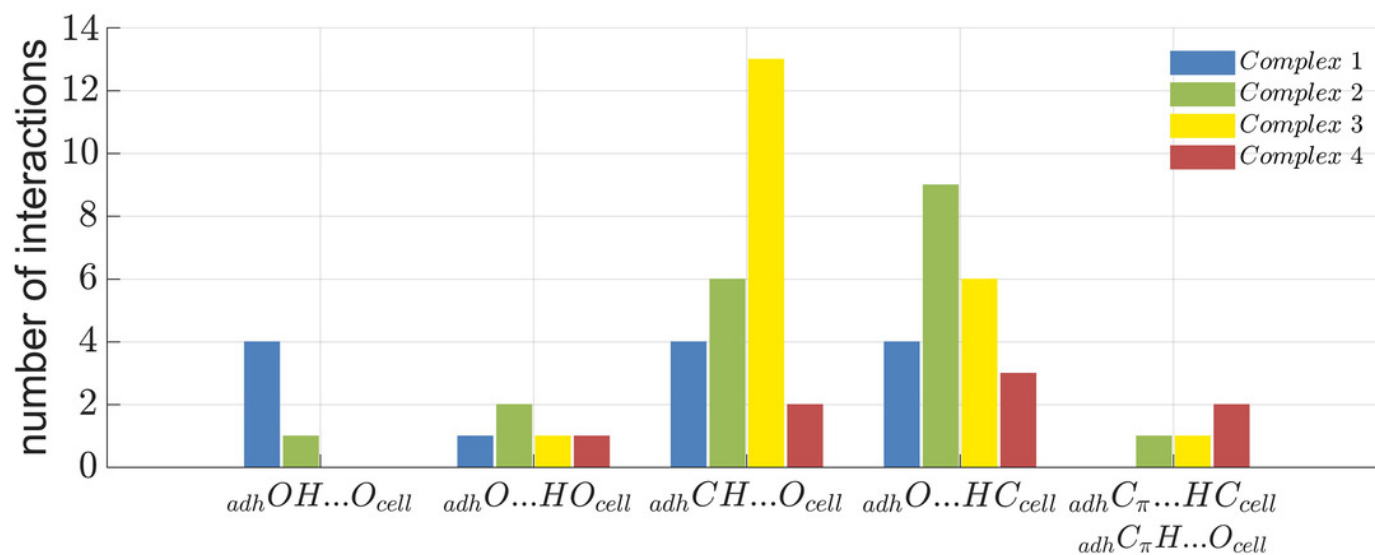




# Figure 7

## Summary of non-covalent interactions

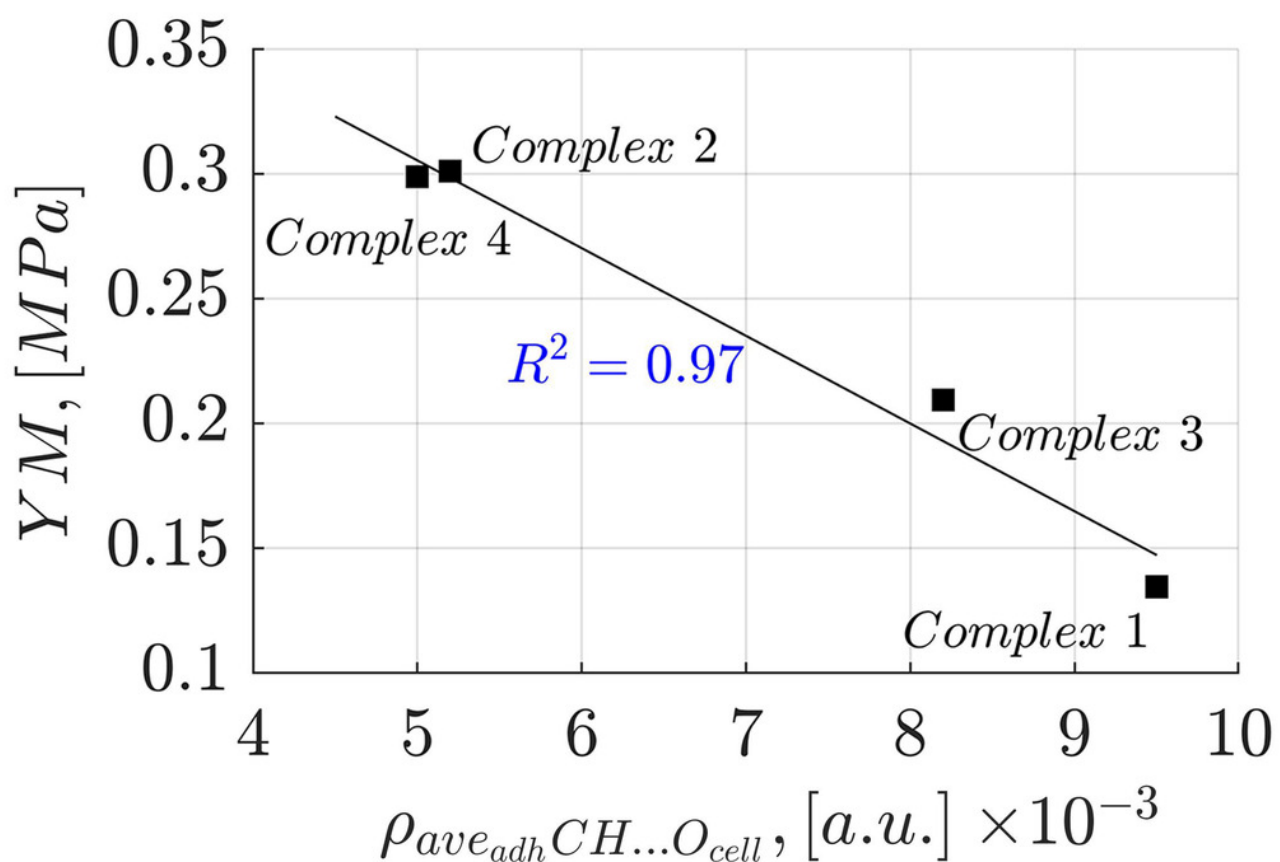
Summary of non-covalent interactions found in the four oligolignol-cellulose complexes.



## Figure 8

Linear correlation between  $_{adh}CH\cdots O_{Cell}$  interactions and Young's modulus

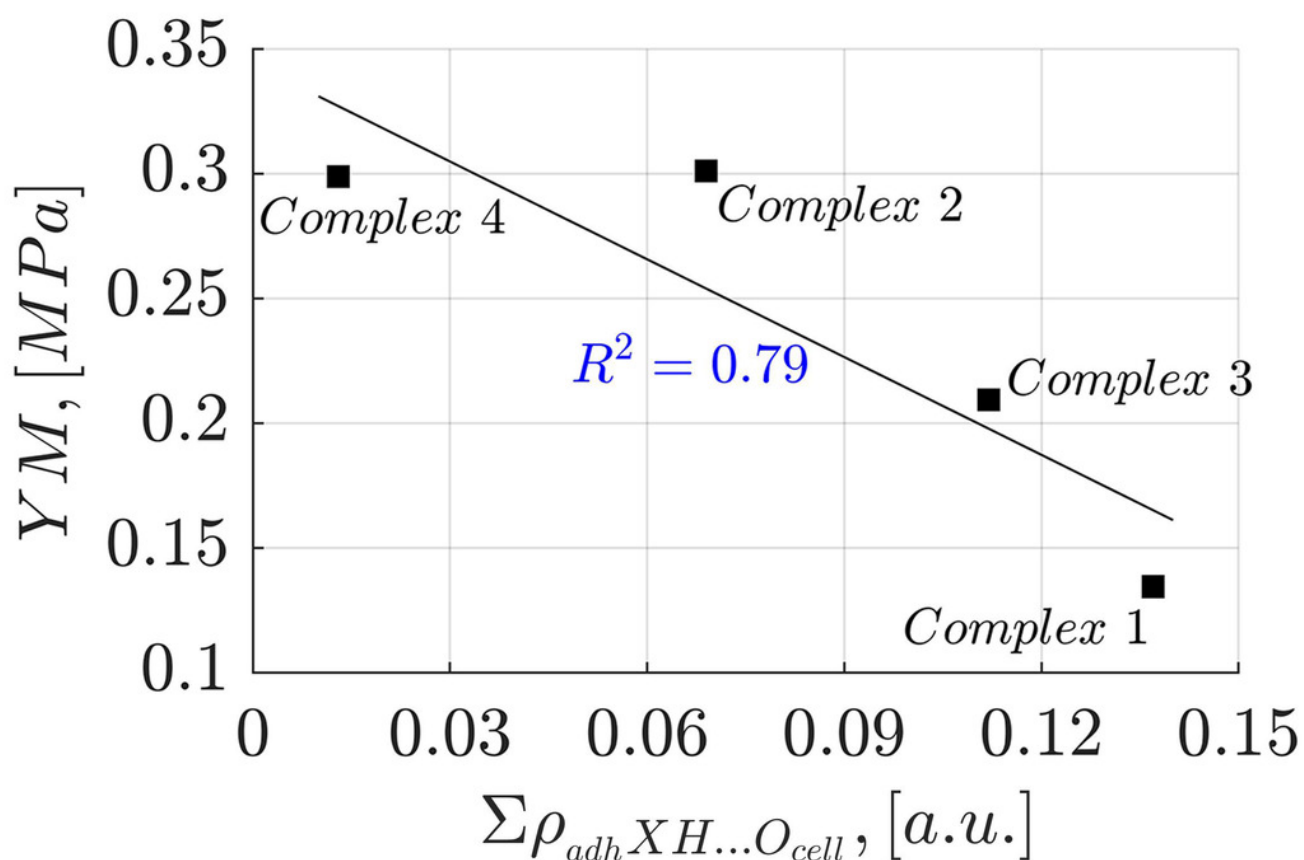
Correlation between the average electron density of the  $_{adh}CH\cdots O_{Cell}$  interactions and Young's modulus for the evaluated complexes.



## Figure 9

Linear correlation between  ${}_{adh}XH\cdots O_{Cell}$  interactions and Young's modulus

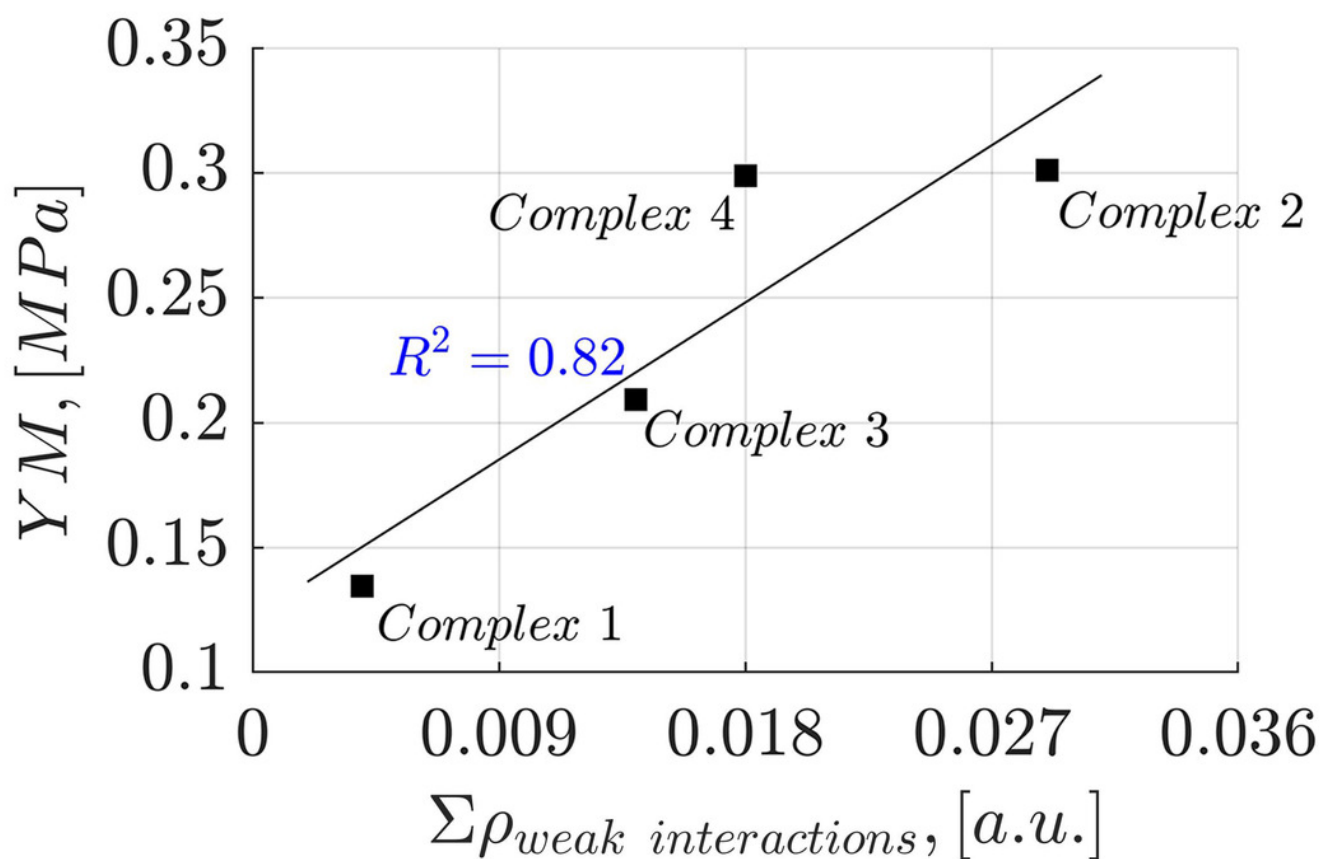
Correlation obtained between the total electron density contribution of  ${}_{adh}XH\cdots O_{Cell}$  interactions and Young's modulus for the evaluated complexes.



## Figure 10

Linear correlation between weak interactions ( $\rho < 0.005$  a.u.) and Young's modulus

Correlation between the total electron density contribution of weak interactions ( $\rho < 0.005$  a.u.) and Young's modulus for the evaluated complexes.



## Figure 11

Correlation the ratio of  ${}_{adh}XH\cdots O_{Cell}$  weak interactions and Young's modulus.

Correlation the ratio of total electron density contribution of  ${}_{adh}XH\cdots O_{Cell}$  and weak interactions and Young's modulus for the evaluated complexes.

